

Cytotoxic compounds from *Goniothalamus repevensis* Pierre ex Finet & Gagnep

Waraporn Chanakul^{a,*}, Vichai Reutrakul^b, Chutima Kuhakarn^b, Samreang Bunteang^b, Pawinee Piyachaturawat^c, Kanoknetr Suksen^c, Sakchai Hongthong^d, Sariyarach Thanasansurapong^b, Narong Nuntasaen^e

^a Faculty of Science, Energy and Environment, King Mongkut's University of Technology North Bangkok, Rayong Campus, Rayong 21120 Thailand

^b Department of Chemistry and Center Excellence for Innovation in Chemistry (PERCH-CIC), Faculty of Science, Mahidol University, Bangkok 10400 Thailand

^c Department of Physiology, Faculty of Science, Mahidol University, Bangkok 10400 Thailand

^d Division of Chemistry, Faculty of Science and Technology, Rajabhat Rajanagarindra University, Chachoengsao 24000 Thailand

^e Department of National Parks, Wildlife and Plant Conservation, Ministry of Natural Resources and Environment, The Forest Herbarium, Bangkok 10900 Thailand

*Corresponding author, e-mail: waraporn.c@sciee.kmutnb.ac.th

Received 31 Aug 2021, Accepted 22 Jan 2022

Available online 15 Apr 2022

ABSTRACT: Phytochemical investigation of the methanol extract of the leaves and twigs of *Goniothalamus repevensis* led to the isolation of five known compounds including two styryl lactones; (–)-goniodiol-7-monoacetate (1) and (–)-goniodiol diacetate (2) as well as three aristolactam alkaloids; piperolactam C (3), aristolactam A III (4), and aristolactam B III (5). The structure determination was established by various spectroscopic methods and by comparison with the literature data. Cytotoxic activity against a panel of mammalian cancerous cell lines of compounds 1–4 was investigated. Piperolactam C (3) was found to be active in all tested cell lines with IC₅₀ values in the range of 3.30–64.6 μM. (–)-Goniodiol-7-monoacetate (1) exhibited cytotoxic activity only against ASK cells with IC₅₀ value of 10.28 μM. Having been isolated as a major component, compound 1 can be considered chemotaxonomic significance for *G. repevensis*.

KEYWORDS: *Goniothalamus repevensis*, cytotoxicity, styryl lactone, goniodiol, aristolactam

INTRODUCTION

Plants in the *Goniothalamus* genus belong to the Annonaceae family with greater than 130 species recorded worldwide [1], of which about 25 species grow in Thailand [2]. Various plants in the *Goniothalamus* genus have long been used in Thai traditional medicines. In Narathiwat Province, Thailand, the flowers of *G. giganteus* and *G. malayanus* have been used as cardiogenic. In the Northeastern part of Thailand including Yasothon, Ubon Ratchathani, and Chaiyaphum Provinces, the flowers of *G. marcanii* and wood of *G. laoticus* have been used for treatment of infectious diseases in children [3].

Various *Goniothalamus* species are known to produce structurally diverse metabolites, comprising acetogenins [4–10], styryl lactones [11–17], alkaloids [14, 16–19], and flavonoids [20]. Several acetogenins and styryl lactones exhibited potent cytotoxic activity against various cancerous cell lines [21]. Some of the alkaloids and flavonoids displayed a broad spectrum of biological activities such as antifungal, antiparasitic, antimycobacterial, insecticidal, antimalarial, anti-inflammatory, and immunosuppressive activities as well as inhibitory activity toward platelet-activating

factor (PAF) receptor binding activity [22]. According to a vast biologically active compounds having been reported, plants in the *Goniothalamus* genus have drawn attention and been considered important natural resources to be searched for their potential metabolites for further development of anti-cancer drug candidates [23].

Goniothalamus repevensis Pierre Finet & Gagnep, called in Thai “Sad-Siam”, is an endemic species and distributes in evergreen forest, 600–900 m above mean sea level in Chanthaburi Province, located in the Southeastern part of Thailand. Notably, small, and pink flowers are the key characteristics of *G. repevensis*, which is distinguishable from other plants in this genus. To the best of our knowledge, no phytochemical investigation or biological study on *G. repevensis* has been reported. Thus, the methanol extract of leaves and twigs of *G. repevensis* was preliminarily screened for cytotoxic activity. The results indicated that it exhibited significant cytotoxic activity against a panel of mammalian cancerous cell lines. In this work, the isolation and structural determination of chemical constituents from the methanol extract of the leaves and twigs of *G. repevensis* were firstly reported. Some isolated compounds were evaluated for their cytotoxic

activity.

MATERIALS AND METHODS

General experimental procedures

Optical rotations were measured on a JASCO DIP-370 digital polarimeter (Japan). UV absorption spectra were measured in MeOH on a Shimadzu 2800 UV-visible spectrophotometer (Japan); principal bands (λ_{\max}) were recorded as wavelengths (nm) and $\log \epsilon$. IR spectra were recorded on a Perkin-Elmer Frontier FT-IR spectrometer (USA); major bands (ν_{\max}) were recorded in wave number (cm^{-1}). High resolution mass spectra (HRMS) were recorded on a Micro-mass model VQ-TOF spectrometer (Waters, USA). ^1H NMR and ^{13}C NMR spectra were recorded on a Bruker DPX-400 or a Bruker Avance-500 spectrometer (Switzerland) in CDCl_3 or $\text{DMSO}-d_6$ using tetramethylsilane (TMS) or residual non-deuterated solvent peak as an internal reference. Chromatographic methods were performed by using silica gel 60 (70–230 mesh, Merck, Germany) for column chromatography, silica gel plates (Silica gel 60 PF₂₅₄, Merck) for preparative TLC, and Sephadex LH 20 (GE Healthcare, USA) for gel filtration. All solvents used for extraction and isolation were distilled at their boiling point ranges prior to use.

Plant materials

The leaves and twigs of *G. repevensis* Pierre ex Finet & Gagnep were collected from Khao Khitchakut National Park, Chanthaburi Province, Thailand in October 2012. The plant was identified by Narong Nuntasaen, and the voucher specimen (BKF 192543) was deposited at the Forest Herbarium, Department of National Parks, Wildlife and Plant Conservation, Bangkok, Thailand.

Extraction and isolation

The air-dried and finely powdered leaves and twigs of *G. repevensis* (1.88 kg) were macerated with MeOH (6 times \times 4 l) at room temperature. After filtration and removal of the solvent under reduced pressure by a rotary evaporator, vacuum desiccation, and freeze-drying, the crude MeOH extract (181.5 g, 9.63%) was obtained. The crude MeOH extract was screened for cytotoxic activity against five mammalian cancerous cell lines, and the results were shown in Table 1. A part of the MeOH extract (135.0 g) was subjected to vacuum column chromatography over silica gel (400 g) and eluted with EtOAc-hexanes and MeOH-EtOAc gradients. Elution was initially conducted with EtOAc-hexanes gradient (0:1, 1:9, 1:4, 3:7, 2:3, 1:1, 7:3, and 1:0 v/v, 2 l each), followed by MeOH-EtOAc gradient (0:1, 1:9, 1:4, 1:1, and 1:0, 2 l each). Fractions (500 ml each) were collected and combined on the basis of their TLC characteristics to afford six fractions (F1–F6). Fraction F2 (14.3 g, eluted with EtOAc-hexanes 1:9, 1:4, and 3:7 v/v) provided

goniodiol-7-monoacetate (1) (530.0 mg, 0.39%) after crystallization from EtOAc-hexanes. The mother liquor of fraction F2 (14.2 g) was subjected to silica gel column chromatography and eluted with EtOAc-hexanes gradient hexanes (5:95, 1:9, 1:4, 3:7, 2:3, and 1:1 v/v, 2 l each) to give 10 subfractions (F2-1 to F2-10). Piperolactam C (3) (40.0 mg, 0.03%), goniodiol diacetate (2) (20.0 mg, 0.01%), and aristolactam B III (5) (21.6 mg, 0.02%) were obtained from subfractions F2-4, F2-7, and F2-8, respectively, after Sephadex LH-20 column chromatography (1:1 v/v $\text{CH}_2\text{Cl}_2/\text{MeOH}$ as eluent). Fraction F3 (15.8 g, eluted with EtOAc-hexanes 2:3 v/v) was subjected to silica gel column chromatography and eluted with EtOAc-hexanes gradient (1:4, 3:7, 2:3, 1:1, and 3:2 v/v, 2 l each) to give six subfractions (F3-1 to F3-6). Aristolactam A III (4) (20.0 mg, 0.01%) was isolated from subfraction F3-6. See Supplementary data for ^1H NMR and ^{13}C NMR data of compounds 1–5 (Tables S1–S5).

Cytotoxic assays

The crude MeOH extract and isolated compounds were evaluated for cytotoxic activity using the standard *in vitro* sulforhodamine B (SRB) assay. This method measures the cellular protein content of adherent and suspension cultures in 96 well microtiter plates. [24, 25]. Ellipticine, a potent cytotoxic plant alkaloid, was used as a positive control. Five cancerous cell lines were employed, including KB (human epidermoid carcinoma in the mouth), HT-29 (human colorectal adenocarcinoma), MCF-7 (human breast carcinoma), A549 (human lung carcinoma), and ASK (rat glioma cells). For noncancerous cell line, Hek293 (noncancerous human embryonic kidney cells) was employed for screening of the MeOH extract while CL (human normal liver hepatic cells) was employed for screening of the pure compounds. The potency of cytotoxic activity is expressed as 50% inhibitory dose (IC_{50}).

RESULTS AND DISCUSSION

The MeOH extract of the leaves and twigs of *G. repevensis* was obtained by maceration extraction. Preliminary screening indicated that the crude MeOH extract showed significant cytotoxic activity against a panel of mammalian cancerous cell lines including KB, HT29, MCF7, A549, ASK, and Hek293 cell lines with IC_{50} values in the range of 1.58–3.89 $\mu\text{g}/\text{ml}$. The isolation and purification of chemical compounds were performed by chromatographic techniques and crystallization to yield five known compounds 1–5. The structures of the isolated compounds were elucidated by spectroscopic techniques including UV, IR, NMR, and MS analysis. By comparison of the spectroscopic data of compounds 1–5 with those previously reported in the literature; see Tables S1–S5, compounds 1–5 were identified to be (–)-goniodiol-7-monoacetate (1) [26], (–)-goniodiol diacetate (2) [26], piperolactam C (3)

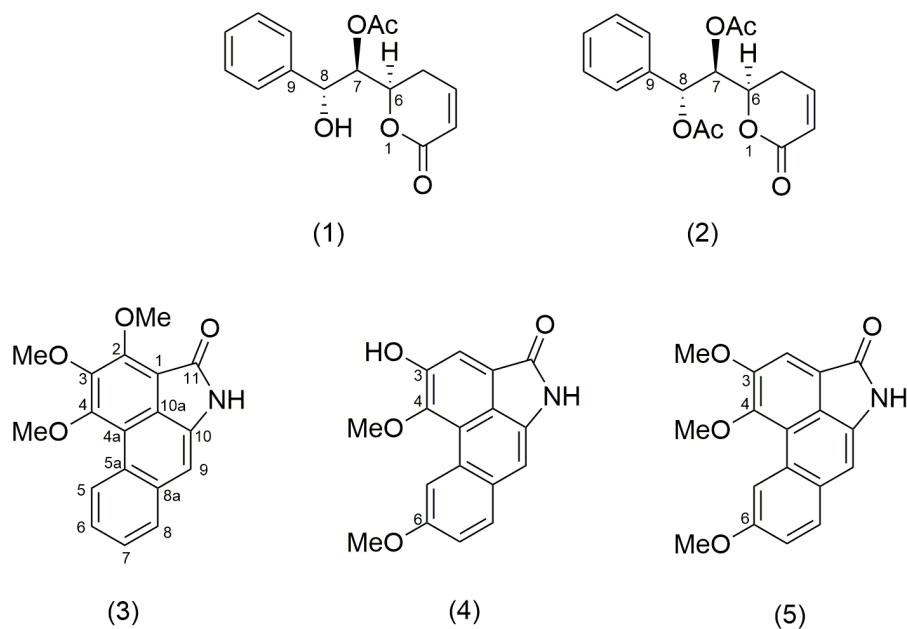


Fig. 1 Isolated compounds 1–5 from leaves and twigs of *G. repevensis*.

Table 1 Cytotoxicity of MeOH extract (IC_{50} , $\mu\text{g/ml}$) and isolated compounds 1–4 (IC_{50} , μM).

Extract/Compound	Cell line (IC_{50}) ^a					
	KB	HT-29	MCF 7	A549	ASK	CL ^d
MeOH extract ^b	1.58	3.81	3.89	3.16	3.17	2.01
1	24.00	35.11	55.59	37.64	10.28	32.39
2	29.06	27.68	35.31	28.30	13.29	26.14
3	8.47	> 64.66	31.20	11.90	3.30	8.21
4	10.23	45.72	63.63	46.06	7.38	41.11
Elliphticine ^c	2.03	2.31	1.95	1.99	1.95	2.03

^a IC_{50} value presented in μM was obtained from three independent experiments for pure compounds. KB, human nasopharyngeal carcinoma; HT-29, human colorectal adenocarcinoma; MCF-7, human breast carcinoma; A549, human lung carcinoma; ASK, rat glioma cells; and CL, human normal liver hepatic cells.

^b IC_{50} presented in $\mu\text{g/ml}$ for extract. ^c Elliphticine was used as a positive control for cytotoxicity assay. ^d Using Hek293, noncancerous human embryonic kidney cells, in testing for the extract.

[27], aristolactam AIII (4), and aristolactam BIII (5) [28, 29]; see Fig. 1. Compounds 1–4 were evaluated for their cytotoxic activity against a panel of mammalian cancerous cell lines as well as noncancerous human liver hepatic cells (CL); see Table 1. All tested compounds were found active against ASK cancer cells with IC_{50} in the range of 3.30–13.29 μM with compound 3 being the most active compound (IC_{50} 3.30 μM). Compound 3 was also active against KB and A549 cancer cells with IC_{50} values of 8.47 and 11.90 μM , respectively. However, compound 3 was toxic to CL cells with IC_{50} value of 8.21 μM . (–)-Goniodiol-7-monoacetate (1), the major component isolated (0.39%), exhibited selective cytotoxic activity against ASK cell line with IC_{50} value of 10.28 μM while it was less toxic to CL (IC_{50} 32.39 μM). Based on these results, compound 1 can be considered a

potential candidate for further in-depth study of its cytotoxic effects.

For the chemotaxonomic significance, styryl lactones are the major types of compounds found in plants of *Gonithalamus* genus. In particular, (–)-goniodiol-7-monoacetate (1), first isolated from the leaves and stems of *G. amuyon*, is commonly found in various plant species belonging to the *Gonithalamus* genus [26, 30, 31]. Recently, it was reported to be isolated from the barks of *G. lanceolatus* [32]. (–)-Goniodiol-7-monoacetate (1) showed potent cytotoxic activity against human epidermoid carcinoma of the nasopharynx (KB), murine lymphocytic leukemia (P-388), human melanoma (RPMI), and central nervous system carcinoma (TE671) with ED_{50} values of < 0.1 $\mu\text{g/ml}$ [31]. Therefore, (–)-goniodiol-7-monoacetate (1) was a potential candidate obtained

from natural resources and deserves comprehensive study toward its cytotoxic effects for further development of anti-cancer drug.

CONCLUSION

The present work described the first phytochemical investigation of *Goniothalamus repevensis*, leading to the isolation of five known compounds including (–)-goniodiol-7-monoacetate (1), (–)-goniodiol diacetate (2), piperolactam C (3), aristolactam AIII (4), and aristolactam BIII (5) from the cytotoxic MeOH extract of the leaves and twigs. The structure determination of the isolated compounds was established by means of spectroscopic methods and by comparison with the literature data. Compounds 1–4 showed cytotoxic activity against a panel of mammalian cancer cell lines with IC₅₀ values in the range of 3.30–64.6 μM with compound 3 displaying significant cytotoxic activity against ASK and KB cell lines with IC₅₀ values of 3.30 and 8.47 μM, respectively. Compound 1 exhibited cytotoxic activity only against ASK cell line with an IC₅₀ value of 10.28 μM while it was less toxic to CL cell lines (human normal liver hepatic cells) with an IC₅₀ value of 32.39 μM. Compound 1, being isolated as a major component, can be potentially considered the chemotaxonomic significance for *G. repevensis* species.

Appendix A. Supplementary data

Supplementary data (Tables S1–S5) associated with this article is available upon request from the corresponding author.

Acknowledgements: This work was supported by the Thailand Research Fund [MRG5980036]. We are grateful to the Center of Excellence for Innovation in Chemistry (PERCH-CIC), Ministry of Higher Education, Science, Research and Innovation, Thailand and Department of Chemistry, Faculty of Science, Mahidol University for laboratory equipment and spectroscopic measurements.

REFERENCES

- Choo C-Y, Abdullah N, Diederich M (2014) Cytotoxic activity and mechanism of action of metabolites from the *Goniothalamus* genus. *Phytochem Rev* **13**, 835–851.
- Saunders RMK, Chalermglin P (2008) A synopsis of *Goniothalamus* species (Annonaceae) in Thailand, with descriptions of three new species. *Bot J Linn Soc* **156**, 355–384.
- Chuakul W, Sornthorncharenon N (2003) Ethnomedical uses of Thai Annonaceous plant (1). *Thai J Phytopharm* **10**, 25–32.
- Fang XP, Anderson JE, Smith DL, McLaughlin JL (1992) Gigantetronenin and gigantrionenin: novel cytotoxic acetogenins from *Goniothalamus giganteus*. *J Nat Prod* **55**, 1655–1663.
- Gu ZM, Fang XP, Zeng L, Song R, Ng JH, Wood KV, Smith DL, McLaughlin JL (1994) Gonionenin a new cytotoxic annonaceous acetogenin from *Goniothalamus giganteus* and the conversion of mono-THF acetogenins to bis-THF acetogenins. *J Org Chem* **59**, 3472–3479.
- Alali FQ, Zhang Y, Rogers L, McLaughlin JL (1997) (2,4-*cis* and *trans*)-Gigantecinone and 4-deoxygigantecin, bioactive nonadjacent bis-tetrahydrofuran Annonaceous acetogenins from *Goniothalamus giganteus*. *J Nat Prod* **60**, 929–933.
- Alali FQ, Rogers L, Zhang Y, McLaughlin JL (1999) Goniotriocin and (2,4-*cis*- and -*trans*)-xyломaticinones, bioactive Annonaceous acetogenins from *Goniothalamus giganteus*. *J Nat Prod* **62**, 31–34.
- Jiang Z, Yu DQ (1997) New type of monotetrahydrofuran ring acetogenins from *Goniothalamus donnaiensis*. *J Nat Prod* **60**, 122–125.
- Jiang Z, Chen Y, Chen RY, Yu DQ (1998) Linear acetogenins from *Goniothalamus donnaiensis*. *Phytochemistry* **49**, 769–775.
- Tantithanaporn S, Wattanapiromsakul C, Itharat A, Keawpradub N (2011) Cytotoxic activity of acetogenins and styryl lactones isolated from *Goniothalamus undulatus* Ridl. root extracts against a lung cancer cell line (COR-L23). *Phytomedicine* **18**, 486–490.
- Fang XP, Anderson JE, Chang CJ, McLaughlin JL (1991) Two new styryl lactones, 9-deoxygoniopyrpyrone and 7-epi-goniofurfurone from *Goniothalamus giganteus*. *J Nat Prod* **54**, 1034–1043.
- Wang S, Zhang YJ, Chen RY, Yu DQ (2002) Gonio-lactones A–F, six new styrylpyrone derivatives from the roots of *Goniothalamus cheliensis*. *J Nat Prod* **65**, 835–841.
- Zhu JX, Yu DL, Huang WH, Sun L, Lv Y, Zheng QT, Lee KH, Yu JG (2012) Goniodilactone and gonioheptenolactone, two novel cytotoxic styryllactones from the leaves of *Goniothalamus cheliensis*. *Chinese Chem Lett* **23**, 583–586.
- Suchaichit N, Kanokmedhakul K, Panthama N, Poopaisit K, Moosophon P, Kanokmedhakul S (2015) A 2H-tetrahydropyran derivative and bioactive constituents from the bark of *Goniothalamus elegans* Ast. *Fitoterapia* **103**, 206–212.
- Sangrueng K, Sanyacharernkul S, Nantapap S, Nantasaen N, Pompimon W (2015) Bioactive Goniothalamine from *Goniothalamus tapis* with cytotoxic potential. *Am J Appl Sci* **12**, 650–653.
- Jaidee W, Andersen RJ, Patrick BO, Pyne SG, Muanprasat C, Borwornpinyo S, Laphookhieo S (2019) Alkaloids and styryllactones from *Goniothalamus cheliensis*. *Phytochemistry* **157**, 8–20.
- Meesakul P, Jaidee W, Richardson C, Andersen RJ, Patrick BO, Willis AC, Muanprasat C, Wang J, et al (2020) Styryllactones from *Goniothalamus tamirensis*. *Phytochemistry* **171**, ID 112248.
- Tip-pyang S, Limpipatwattana Y, Khumkratok S, Siripong P, Sichaem J (2010) A new cytotoxic 1-azaanthraquinone from the stems of *Goniothalamus laoticus*. *Fitoterapia* **81**, 894–896.
- Lekphrom R, Kanokmedhakul S, Kanokmedhakul K (2009) Bioactive styryllactones and alkaloid from flowers of *Goniothalamus laoticus*. *J Ethnopharm* **125**, 47–50.
- Banjerpongchai R, Khaw-on P, Pompimon W (2016) Phytochemicals from *Goniothalamus griffithii* induce human cancer cell apoptosis. *Asian Pac J Cancer Prev* **17**, 3281–3287.
- Wiat C (2007) *Goniothalamus* species: A source of drugs

- for the treatment of cancers and bacterial infections, *Evid-based Complement Altern Med* **4**, ID 635095.
22. Seyed MA, Jantan I, Bukhari SNA (2014) Emerging anticancer potentials of goniothalamine and its molecular mechanisms. *BioMed Res Int* **2014**, ID 536508.
 23. Tran DT, Mai HDT, Pham VC, Nguyen VH, Guéritte F, Nguyen QV, Tran TA, Chau VM (2013) Alkaloids and styryllactones from the leaves of *Goniothalamus tamirensis*. *Phytochem Lett* **6**, 79–83.
 24. Likhitwittayawuid K, Angerhofer CK, Cordell GA, Pezuto JM, Ruangrunsi N (1993) Cytotoxic and anti-malarial bisbenzylisoquinoline alkaloids from *Stephania erecta*. *J Nat Prod* **56**, 30–38.
 25. Skehan P, Storeng R, Scudiero D, Monks A, McMahon J, Vistica D, Warren JT, Bokesch H, et al (1990) New colorimetric cytotoxicity assay for anticancer-drug screening. *J Natl Cancer Inst* **82**, 1107–1112.
 26. Wu YC, Chang FR, Duh CY, Wang SK, Wu TS (1992) Cytotoxic styrylpyrones of *Goniothalamus amuyon*. *Phytochemistry* **31**, 2851–2853.
 27. Desai SJ, Prabhu BR, Mulchandani NB (1988) Aristolactams and 4,5-dioxoaporphines from *Piper longum*. *Phytochemistry* **27**, 1511–1515.
 28. Priestap HA (1985) Seven aristolactams from *Aristolochia argentina*. *Phytochemistry* **24**, 849–852.
 29. Mix DB, Guinaudeau H, Shamma M (1982) The aristolochic acids and aristolactams. *J Nat Prod* **45**, 657–666.
 30. Lan YH, Chang FR, Yang YL, Wu YC (2006) New constituents from stems of *Goniothalamus amuyon*. *Chem Pharm Bull* **54**, 1040–1043.
 31. Wu YC, Duh CY, Chang FR, Chang GY, Wang SK, Chang JJ, Mcphail DR, Mcphail AT, et al (1991) The crystal structure and cytotoxicity of goniodiol-7-monoacetate from *Goniothalamus amuyon*. *J Nat Prod* **54**, 1077–1081.
 32. Rasol NE, Ahmad FB, Mai CW, Bihud NV, Abdullah F, Awang K, Ismail NH (2018) Styryl lactones from roots and barks *Goniothalamus lanceolatus*. *Nat Prod Commu* **13**, 1575–1578.