

Dietary flavonoids against various breast cancer subtypes: a molecular docking study

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ABSTRACT: Breast cancer is female most frequent diagnosed cancer and the leading cause of cancer death. The consumption of dietary flavonoids is reported to cause significant breast cancer risk reduction. In vitro studies often used aglycone flavonoids rather than its conjugated form that actually present in human body. Thus its mechanism against breast cancer has not been elucidated completely. The present study aimed to investigate the possible mechanism of dietary flavonoids against breast cancer by in silico study. Conjugated flavonoids were docked to ER (estrogen receptor), HER2 (human epidermal growth factor receptor 2) and EGFR (epidermal growth factor receptor) kinase domains. The molecular docking of 22 flavonoid conjugates towards EGFR and HER2 kinase domain, and ER was successfully performed. Potential binders to proteins: epicatechin conjugates to ER (−8.7 kcal/mol), isoflavone conjugates to HER2 kinase domain (−10.7 kcal/mol), and epigallocatechin and epicatechin conjugates to EGFR kinase domain (−9.2 kcal/mol), were suggested. Supported by other studies, conjugated flavonoids may exert similar inhibitory and agonistic properties to their parent flavonoids. Taken together, the present study showed possible effects of dietary flavonoids against various breast cancer subtypes.

KEYWORDS: molecular docking simulation, diet, flavonoids, antineoplastic agents, breast neoplasms

INTRODUCTION

According to GLOBOCAN 2018 database, breast cancer is female most diagnosed cancer and the leading cause of cancer death¹. Breast cancer is heterogeneous disease, comprised of various subtypes observable by the presence of the predictive molecular markers. Breast cancer can be categorized into: Luminal A (ER⁺, PR^{+/-}, HER⁻), Luminal B (ER⁺, PR^{+/-}, HER2⁺), HER2 (ER⁻, PR⁻, HER2⁺), basal like and claudin low (triple negatives)². Each has a different prognosis and responds differently to cancer treatment. Luminal A has the best prognosis, while HER2⁺ and triple negative breast cancer (TNBC) have the poorest³. Today breast cancer endorsed to be treated with endocrine therapy, targeted therapy, and cytotoxic chemotherapy⁴. Breast cancer with high expression of estrogen receptor

(ER) and progesterone receptor (PR) is sensitive against endocrine therapy. ER inhibition treatment in Luminal A and B breast cancer was proven to be effective and safe⁵. While HER2⁺ breast cancer was effectively treated using targeted therapy with trastuzumab or lapatinib⁶. But different from previous subtypes, triple negative breast cancer is not responded very well to hormone treatment and HER2 antibody, and often treated with systemic chemotherapy. Previous study found that epidermal growth factor receptor (EGFR) kinase inhibitor, gefitinib, was able to halt the TNBC cell outgrowth in vitro⁷. Thus ER, HER2, EGFR served as important targets in breast cancer treatment. It is also possible other chemical compounds found in food may also interacts with these particular proteins. Flavonoid is the most common phytochemical compound found

ubiquitously in human diet^{8,9} and has huge impact in human health. In vitro studies showed that flavonoids have wide range of biological activity antioxidant, anti-inflammatory, anti-microbial, anti-fungal, antiviral, and anti-cancer^{9,10}. Consumption of flavonoids is related with less risk of cardiovascular diseases and stroke^{11,12}. Other studies found that the intake of flavonoids improved the outcome of the gastric and lung cancer^{13,14}. Human study on consumption of food rich in flavonoids, green tea, against breast cancer showed mixed results. Case studies has shown green tea intake was correlated with significant breast cancer risk reduction¹⁵⁻¹⁷, while recent prospective cohort studies showed no correlation¹⁸. Soy products were rich in isoflavone and high soy intake was modestly associated with reduced breast cancer risk¹⁹.

It is important to note that flavonoid is quickly metabolized in the body. After ingested, glycoside flavonoid found in plant materials is subjected to deglycosylation, releasing aglycone compound that readily absorbed by the intestine lining²⁰. Once entered circulatory system, flavonoid is immediately transported to liver and undergoes extensive metabolism. Phase metabolism II transformed free aglycone onto flavonoid conjugates by adding glucuronides and sulphate moiety²¹. Because of this, aglycone flavonoids are rarely found in plasma. Previous in vitro studies often used aglycone flavonoids rather than its conjugated forms that present in human body. Thus its mechanism against breast cancer has not been elucidated completely, since conjugation may affect how the molecules behave. To address this, conjugated flavonoids found in plasma after ingesting food rich in flavonoid was subjected to molecular docking against ER, HER2, EGFR. The present study describes possible mechanism how the dietary flavonoids may contribute against breast cancer.

MATERIALS AND METHODS

Molecular docking towards EGFR, HER2, and ER

Conjugated flavonoids found in plasma after ingesting food rich in flavonoids as previously reported from other studies were used²²⁻²⁴. Structural data of conjugated flavonoids were retrieved from PubChem database (Fig. 1) (pubchem.ncbi.nlm.nih.gov). Co-crystallized structures of ER-4-hydroxytamoxifen (PDB ID: 3ERT)²⁵, HER2-SYR127063 (PDB ID: 3PP0)²⁶, and EGFR-gefitinib (PDB ID: 4WKQ) were obtained from RCSB database (rcsb.org). Crystal structure data were

Table 1 Predicted binding affinity (kcal/mol) of flavonoid conjugates and known inhibitors towards ER, HER2, and EGFR.

Flavonoid conjugate	Binding affinity		
	ER	HER2	EGFR
Gefitinib	-	-	-8.8
SYR127063	-	-11.0	-
4-hydroxytamoxifen	-9.7	-	-
(-)-Epicatechin-3-gallate	-8.7	-8.5	-8.6
(-)-Epigallocatechin-3-gallate	-7.0	-8.6	-9.2
(-)-Epigallocatechin-7-gallate	-7.4	-8.9	-8.0
4'-Methylepicatechin-5-sulfate	-7.2	-8.8	-7.7
4'-Methylepicatechin-7-sulfate	-7.9	-9.6	-7.2
4'-Methyl-epigallocatechin-3'-glucuronide	-7.9	-9.9	-9.1
4'-Methyl-epigallocatechin-7-glucuronide	-6.5	-9.0	-8.2
Daidzein-4'-sulfate	-7.1	-9.4	-8.5
Daidzein-7-sulfate	-6.6	-9.2	-8.4
Epicatechin-3'-glucuronide	-8.5	-9.5	-9.1
Epicatechin-3'-sulfate	-7.7	-9.1	-8.2
Epicatechin-5-sulfate	-7.1	-8.6	-8.4
Epicatechin-7-glucuronide	-7.5	-9.3	-8.3
Epigallocatechin-3'-glucuronide	-7.7	-9.7	-9.1
Epigallocatechin-7-glucuronide	-7.2	-8.3	-8.3
Genistein-4'-O-glucuronide	-8.3	-10.7	-8.0
Genistein-4'-sulfate	-7.3	-9.7	-8.3
Genistein-7-O-glucuronide	-6.8	-9.4	-7.7
Genistein-7-sulfate	-6.4	-9.1	-8.6
Isorhamnetin-3-O-glucuronide	-7.6	-8.3	-8.8
Quercetin-3'-glucuronide	-8.7	-8.8	-8.7
Quercetin-3'-sulfate	-8.1	-9.5	-8.4

prepared by removing solvent and extracting bound ligand. AutoDock vina was used in molecular docking under default settings. The docking methodology was validated by redocking the extracted bound ligand. Chimera was used on visualization in this study. Intramolecular analysis was performed using Pose View, available at Protein Plus (proteins.plus)²⁷.

RESULTS

Molecular docking analysis

Redocking was performed to evaluate software and docking parameters used. The root mean square deviation between docked and crystal compounds was less than 2Å except for EGFR bound ligand (gefitinib). This is due to the 6-propylmorpholino moiety of gefitinib sticking out to solvent and able to move freely²⁸. Thus, AutoDock Vina has favourable accuracy and proceeds the docking of flavonoid-

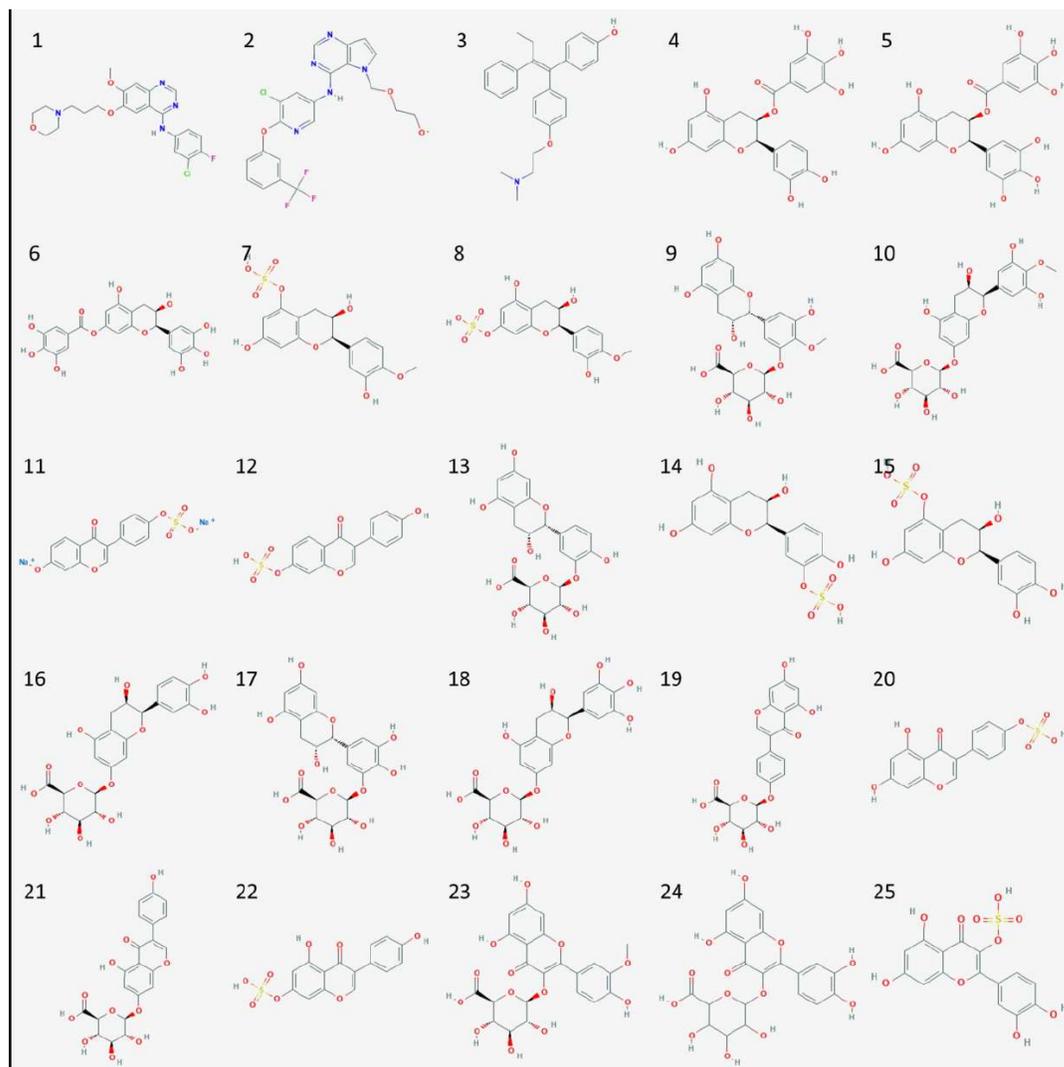


Fig. 1 The 2 dimensional structure of all studied ligands: (1) gefitinib, (2) SYR127063, (3) 4-hydroxytamoxifen, (4) (-)-epicatechin-3-gallate, (5) (-)-epigallocatechin-3-gallate, (6) (-)-epigallocatechin-7-gallate, (7) 4'-methylepicatechin-5-sulfate, (8) 4'-methylepicatechin-7-sulfate, (9) 4'-methyl-epigallocatechin-3'-glucuronide, (10) 4'-methyl-epigallocatechin-7-glucuronide, (11) daidzein-4'-sulfate, (12) daidzein-7-sulfate, (13) epicatechin-3'-glucuronide, (14) epicatechin-3'-sulfate, (15) epicatechin-5-sulfate, (16) epicatechin-7-glucuronide, (17) epigallocatechin-3'-glucuronide, (18) epigallocatechin-7-glucuronide, (19) genistein-4'-O-glucuronide, (20) genistein-4'-sulfate, (21) genistein-7-O-glucuronide, (22) genistein-7-sulfate, (23) isorhamnetin-3-O-glucuronide, (24) quercetin-3'-glucuronide, (25) quercetin-3'-sulfate.

conjugates. The molecular docking was performed to assess possible binding conformation of flavonoid conjugates towards receptors and possible biological actions of these compounds. The molecular docking of 22 flavonoid conjugates towards EGFR and HER2 kinase domain, and ER was successfully performed. The predicted binding affinity value of flavonoid conjugates was compared to each other and receptor bound ligand (Table 1).

In the present study, based on molecular docking, epicatechin and quercetin conjugates ((-)-epicatechin-3-gallate, quercetin-3'-glucuronide, and epicatechin-3'-glucuronide) were predicted as a potential binder towards estrogen receptor. HER2 kinase domain was predicted to interact strongly towards genistein and epigallocatechin conjugates (genistein-4'-O-glucuronide, genistein-4'-sulfate, epigallocatechin-3'-glucuronide,

Table 2 Hydrogen bond formed by flavonoids conjugates and known inhibitors towards EGFR, HER2, and ER.

Flavonoid conjugate	Receptor	Hydrogen bond
4-hydroxytamoxifen	ER	Asp394 Glu353
(-)-Epicatechin-3-gallate		Leu346 Thr347 Asp351
Quercetin-3'-glucuronide		Leu387 Glu419 Gly420 Asp351 Glu353
SYR127063 Genistein-4'-O-glucuronide	HER2	Met801 Asp863 Ser783 Thr798
Gefitinib (-)-Epigallocatechin-3-gallate	EGFR	Met793 Glu762 Leu788 Met793 Arg841

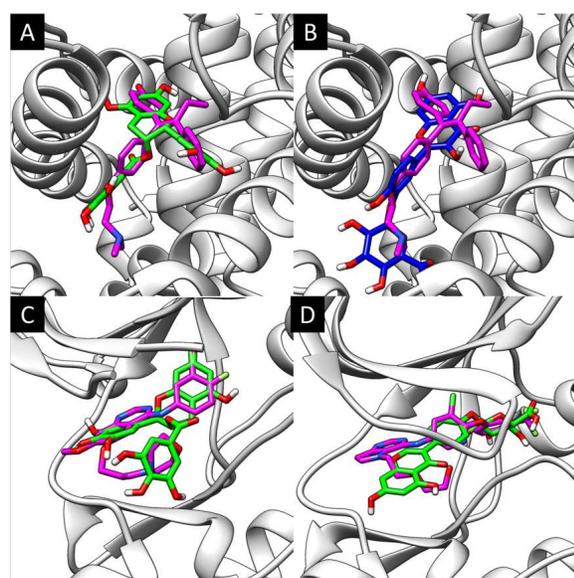


Fig. 2 Superimposed binding mode between flavonoid conjugate and known inhibitor towards ER, HER2, EGFR. (A) and (B) the binding mode of (-)-epicatechin-3-gallate (green), quercetin-3'-glucuronide (blue), and 4-hydroxytamoxifen towards (magenta) ER (light gray). (C) the binding mode of genistein-4'-O-glucuronide (green) and SYR127063 (magenta) towards EGFR kinase domain (light gray). (D) the binding mode of (-)-epigallocatechin-3-gallate (green) and gefitinib (magenta) towards HER2 kinase domain (light gray). The protein represented as ribbon, the compound as stick, and heteroatom represented in a different color from the carbon atom.

and 4'-methyl-epigallocatechin-3'-glucuronide) compared to other compounds. While several catechin derivatives ((-)-epigallocatechin-3-gallate, 4'-methyl-epigallocatechin-3'-glucuronide, epicatechin-3'-glucuronide, and epigallocatechin-3'-glucuronide) were predicted with a high affinity toward EGFR kinase domain as these compounds

surpassed known inhibitor binding score.

Binding mode of the compound with the highest predicted binding affinity was visualized and superimposed with the receptor known inhibitor (Fig. 2). All compounds were found to occupy the active site of the protein. (-)-Epigallocatechin-3-gallate resembled similar binding mode with gefitinib, whereas its core structure aligned with the quinazoline and aniline moiety. While genistein-4'-O-glucuronide, (-)-epicatechin-3-gallate, quercetin-3'-glucuronide only shared small geometrical similarity when compared with the bound ligand. Furthermore, hydrogen bond inferred by Pose View was compared and presented in (Table 2). Similar to gefitinib, (-)-epigallocatechin-3-gallate formed hydrogen bond with Met793. SYR127063 and genistein-4'-O-glucuronide shared no similar intermolecular interaction. Quercetin-3'-glucuronide had similar interaction with 4-hydroxytamoxifen at Glu353. (-)-Epicatechin-3-gallate had the most hydrogen bond towards ESR1.

DISCUSSION

In present study, predicted binding affinity and binding mode of conjugated flavonoids present in plasma after ingestion of dietary flavonoids against ER, EGFR kinase domain, and HER2 kinase domain were characterized *in silico*. The result shows that most compounds with predicted high binding affinity were glucuronide flavonoid conjugates. It is interesting to point out that the predominant flavonoid metabolite found in plasma after an hour ingestion of radiolabelled epicatechin was its glucuronide conjugates²⁹. Thus, the potential compounds found in this study were likely exist in large concentration in plasma after consumption of dietary flavonoids.

More than 70% diagnosed breast cancer was the overexpressed ER³. ER plays important role in development and progression of breast cancer, since ER drives proliferation of mammary cells upon binding with estrogenic hormone³⁰. ER⁺ breast cancer is sensitive against endocrine therapy and is effectively treated using selective estrogen receptor modulators such as tamoxifen³¹. This study found that (-)-epicatechin-3-gallate was a potential inhibitor of ER, because of its high predicted binding affinity and similar binding mode when compared to active metabolite of tamoxifen (4-hydroxytamoxifen). Tamoxifen interacts with several amino acid residues inside the binding pocket, including Leu346, Thr347 and Leu387, forming a van der Waals interaction that stabilize the com-

plex²⁵. Epicatechin conjugates was predicted to be interacted with similar manner. This finding confirmed by other studies whereas epicatechin gallate was able to hamper ER activity through direct inhibition^{32,33}. Quercetin conjugates also have high predicted binding affinity towards ER. But quercetin conjugates may act as an agonist rather than antagonist, since previous study found that aglycone quercetin induced cell proliferation of ER-positive breast cancer cell line through ER stimulation³⁴.

HER2 is a receptor tyrosine kinase which is over-expressed in 30% human breast cancer³. HER2⁺ breast cancer characterized by its aggressive phenotype: high tumorigenicity and invasiveness³⁵. The treatment involved is either by targeting the extracellular domain using trastuzumab or its kinase domain using lapatinib^{4,36}. Previous study showed that flavonoid compounds were able to inhibit human kinases³⁷. Molecular docking study reported that genistein-4'-O-glucuronide had predicted binding affinity close to SYR127063. SYR127063 itself is a potent HER2 kinase domain inhibitor at IC50 of 11 nM²⁶. From the experimental study, genistein was able to attenuate HER2 phosphorylation in BT474 cell line through tyrosine kinase inhibition, thus supported present finding³⁸.

TNBC occurs approximately 10% in breast cancer cases³. TNBC is biologically aggressive and has the poorest prognosis when compared to other subtypes³. Previous study reported that EGFR is a potential target for TNBC⁷. In this work, epigallocatechin and epicatechin metabolites had notable predicted binding affinity towards EGFR kinase domain. This finding supported by another study where epigallocatechin-3-gallate was able to inhibit EGFR activity³⁹. Inhibition of EGFR by epigallocatechin conjugate may also affect HER2 activation, since both proteins are able to form heterodimeric complex and activate each other.

CONCLUSION

Binding affinity and binding mode between conjugated flavonoids found in plasma against ER, HER2, EGFR had been characterized in silico. Supported by other studies, conjugated flavonoids may exert similar inhibitory and agonistic properties to their parent flavonoids. Our study thus confirm and offer possible explanations how dietary flavonoids act against various breast cancer subtypes.

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REFERENCES

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A (2008) Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* **68**, 394–424.
2. Holliday DL, Speirs V (2011) Choosing the right cell line for breast cancer research. *Breast Cancer Res* **13**, ID 215.
3. Hennigs A, Riedel F, Gondos A, Sinn P, Schirmacher P, Marmé F, Jäger D, Kauczor HU, et al (2016) Prognosis of breast cancer molecular subtypes in routine clinical care: A large prospective cohort study. *BMC Cancer* **16**, ID 734.
4. Goldhirsch A, Winer EP, Coates AS, Gelber RD, Piccart-Gebhart M, Thürlimann B, Senn HJ, Panel members, et al (2013) Personalizing the treatment of women with early breast cancer: highlights of the st gallen international expert consensus on the primary therapy of early breast cancer. *Ann Oncol* **24**, 2206–2223.
5. Hwang KT, Kim EK, Jung SH, Lee ES, Kim SI, Lee S, Park HK, Kim J, et al (2018) Tamoxifen therapy improves overall survival in luminal a subtype of ductal carcinoma in Situ: a study based on nationwide korean breast cancer registry database. *Breast Cancer Res Treat* **169**, 11–22.
6. Goldhirsch A, Gelber RD, Piccart-Gebhart MJ, De Azambuja E, Procter M, Suter TM, Jackisch C, Cameron D, et al (2013) 2 years versus 1 year of adjuvant trastuzumab for HER2-positive breast cancer (HERA): An open-label, randomised controlled trial. *Lancet* **382**, 1021–1028.
7. Savage P, Blanchet-Cohen A, Revil T, Badescu D, Saleh SM, Wang YC, Zuo D, Liu L, et al (2017) A targetable EGFR-dependent tumor-initiating program in breast cancer. *Cell Rep* **21**, 1140–1149.
8. Chun OK, Chung SJ, Song WO (2007) Estimated dietary flavonoid intake and major food sources of U.S. adults. *J Nutr* **137**, 1244–1252.
9. Lotito SB, Frei B (2006) Consumption of flavonoid-rich foods and increased plasma antioxidant capacity in humans: cause, consequence, or epiphenomenon? *Free Radic Biol Med* **41**, 1727–1746.
10. Hoensch HP, Oertel R (2015) The value of flavonoids for the human nutrition: short review and Perspectives. *Clin Nutr Exp* **3**, 8–14.

11. Tangney CC, Rasmussen HE (2013) Polyphenols, inflammation, and cardiovascular disease. *Curr Atheroscler Rep* **15**, ID 324.
12. Hollman PCH, Geelen A, Kromhout D (2010) Dietary flavonol intake may lower stroke risk in men and women. *J Nutr* **140**, 600–604.
13. González CA, Sala N, Rokkas T (2013) Gastric cancer: epidemiologic aspects. *Helicobacter* **18**, 34–38.
14. Woo HD, Kim J (2013) Dietary flavonoid intake and smoking-related cancer risk: a meta-analysis. *PLoS One* **8**, ID e75604.
15. Shrubsole MJ, Lu W, Chen Z, Shu XO, Zheng Y, Dai Q, Cai Q, Gu K, et al (2009) Drinking green tea modestly reduces breast cancer risk. *J Nutr* **139**, 310–316.
16. Wu AH, Yu MC, Tseng C-C, Hankin J, Pike MC (2003) Green tea and risk of breast cancer in asian americans. *Int J Cancer* **106**, 574–579.
17. Zhang M, Holman CDJ, Huang J-P, Xie X (2006) Green tea and the prevention of breast cancer: a case-control study in southeast china. *Carcinogenesis* **28**, 1074–1078.
18. Iwasaki M, Inoue M, Sasazuki S, Sawada N, Yamaji T, Shimazu T, Willett WC, Tsugane S (2010) Green tea drinking and subsequent risk of breast cancer in a population to based cohort of Japanese women. *Breast Cancer Res* **12**, ID R88.
19. Wu AH, Yu MC, Tseng C-C, Pike MC (2007) Body size, hormone therapy and risk of breast cancer in asian-american women. *Int J Cancer* **120**, 844–852.
20. Day AJ, Canada FJ, Diaz JC, Kroon PA, Mclauchlan R, Faulds CB, Plumb GW, Morgan MR, et al (2000) Dietary flavonoid and isoflavone glycosides are hydrolysed by the lactase site of lactase phlorizin hydrolase. *FEBS Lett* **468**, 166–170.
21. Crozier A, Del Rio D, Clifford MN (2010) Bioavailability of dietary flavonoids and phenolic compounds. *Mol Aspects Med* **31**, 446–467.
22. Mullen W, Edwards CA, Crozier A (2006) Absorption, excretion and metabolite profiling of methyl, glucuronyl, glucosyl, and sulpho-conjugates of quercetin in human plasma and urine after ingestion of onions. *Br J Nutr* **96**, ID 107.
23. Del Rio D, Calani L, Cordero C, Salvatore S, Pellegrini N, Brighenti F (2010) Bioavailability and catabolism of green tea flavan-3-ols in humans. *Nutrition* **26**, 1110–1116.
24. Hosoda K, Furuta T, Yokokawa A, Ogura K, Hiratsuka A, Ishii K (2008) Plasma profiling of intact isoflavone metabolites by high-performance liquid chromatography and mass spectrometric identification of flavone glycosides daidzin and genistin in human plasma after administration of kinako. *Drug Metab Dispos* **36**, 1485–1495.
25. Shiau AK, Barstad D, Loria PM, Cheng L, Kushner PJ, Agard DA, Greene GL (1998) The structural basis of estrogen receptor/coactivator recognition and the antagonism of this interaction by tamoxifen. *Cell* **95**, 927–937.
26. Aertgeerts K, Skene R, Yano J, Sang BC, Zou H, Snell G, Jennings A, Iwamoto K et al (2011) Structural analysis of the mechanism of inhibition and allosteric activation of the kinase domain of HER2 protein. *J Biol Chem* **286**, 18756–18765.
27. Stierand K, Rarey M (2010) Drawing the PDB: protein-ligand complexes in two dimensions. *ACS Med Chem Lett* **1**, 540–545.
28. Yun CH, Boggon TJ, Li Y, Woo MS, Greulich H, Meyerson M, Eck MJ (2007) Structures of lung cancer-derived EGFR mutants and inhibitor complexes: mechanism of activation and insights into differential inhibitor sensitivity. *Cancer Cell* **11**, 217–227.
29. Ottaviani JI, Borges G, Momma TY, Spencer JP, Keen CL, Crozier A, Schroeter H (2016) The metabolome of [2-14C]-epicatechin in humans: implications for the assessment of efficacy, safety and mechanisms of action of polyphenolic bioactives. *Sci Rep* **6**, ID 29034.
30. Sommer S, Fuqua SA (2001) Estrogen receptor and breast cancer. *Semin Cancer Biol* **11**, 339–352.
31. Narod S, Nazarali S (2014) Tamoxifen for women at high risk of breast cancer. *Breast Cancer Targets Ther* **6**, ID 29.
32. Kuruto-Niwa R, Inoue S, Ogawa S, Muramatsu M, Nozawa R (2000) Effects of tea catechins on the ERE-regulated estrogenic activity. *J Agric Food Chem* **48**, 6355–6361.
33. Goodin MG (2002) Estrogen receptor-mediated actions of polyphenolic catechins in vivo and in vitro. *Toxicol Sci* **69**, 354–361.
34. van der Woude H, ter Veld MG, Jacobs N, van der Saag PT, Murk AJ, Rietjens IM (2005) The stimulation of cell proliferation by quercetin is mediated by the estrogen receptor. *Mol Nutr Food Res* **49**, 763–771.
35. Moasser MM (2007) The oncogene HER2: Its signaling and transforming functions and its role in human cancer pathogenesis. *Oncogene* **26**, 6469–6487.
36. Schroeder R, Stevens C, Sridhar J (2014) Small molecule tyrosine kinase inhibitors of ErbB2/HER2/Neu in the treatment of aggressive breast cancer. *Molecules* **19**, 15196–15212.
37. Williams RJ, Spencer JP, Rice-Evans C (2004) Flavonoids: antioxidants or signalling molecules? *Free Radic Biol Med* **36**, 838–849.
38. Sakla MS, Shenouda NS, Ansell PJ, MacDonald RS, Lubahn DB (2007) Genistein affects HER2 protein concentration, activation, and promoter regulation in BT-474 human breast cancer cells. *Endocrine* **32**, 69–78.
39. Masuda M, Suzui M, Lim JTE, Weinstein IB (2003) Epigallocatechin-3-gallate inhibits activation of HER2/neu and downstream signaling pathways in human head and neck and breast carcinoma cells. *Clin Cancer Res* **9**, 3486–3491.